# Transition Metal-Free One-Pot Synthesis of Fused 1,4-Thiazepin-5(4H)‑ones and Theoretical Study of the S−N Type Smiles Rearrangement Process

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## **S** Supporting Information



ABSTRACT: A series of 1,4-thiazepin-5(4H)-one derivatives were synthesized via a transition metal-free one-pot Smiles rearrangement process at room temperature. Regioselective seven-membered heterocycles were constructed in good to excellent yields. To gain an in-depth understanding of the S−N type Smiles rearrangement mechanism, a theoretical study was also performed by quantum chemistry calculations.

## **ENTRODUCTION**

1,4-Thiazepin-5(4H)-one derivatives have attracted a great deal of interest because of their outstanding biological and medicinal properties. Compounds containing thiazepine functionalities have been broadly used to cure all kinds of diseases. For example, clentiazem is an important calcium channel blocker, used in the treatment of angina pectoris and hypertension.<sup>1</sup> Clotiapine acts as an antipsychotic for the treatment of bipolar disorder, depressive disorder, and schizophrenia.<sup>2</sup> Ethyl 4-(1[1](#page-7-0) propyldibenzo $[b, f]$ [1,4]thiazepine-2-carboxamido)piperidine-1-carboxylate  $(1)$  was reported as a potential in[ve](#page-7-0)rse agonist.<sup>3</sup> N-Hydroxy-6-(11-oxodibenzo $[b, f][1, 4]$  thiazepin-10(11H)-yl)hexanamide (2) and its analogues exhibit high antitum[or](#page-7-0) activities (Scheme 1).<sup>4</sup> Fused thiazepine scaffolds also have antimicrobial,<sup>5</sup> anti-inflammatory,<sup>6</sup> antihypertensive,<sup>7</sup> and analgesic activities.<sup>8</sup>

As a resul[t,](#page-7-0) vari[ou](#page-1-0)s methods [h](#page-7-0)ave been explor[ed](#page-8-0) for the synthesis of  $1,4$ -thiazepin-5( $4H$ )-one scaffolds, including the Friedel−Crafts reaction beginning with 4-chlorobenzenethiol and 2-chloronicotinic acid, $9$  the heating of 4,6-dichloropyrimidin-5-amine with benzenethiols,<sup>10</sup> and the copper-catalyzed Ullmann arylations using 2-[m](#page-8-0)ercaptobenzoic acid and 1-bromo4-methoxy-2-nitrobenzene.<sup>11</sup> Otte[sen](#page-8-0) also reported the synthesis of 1,4-thiazepin-5(4H)-one derivatives by coupling 4chloro-1-fluoro-2-nitroben[zen](#page-8-0)e with 2-mercaptobenzoate,<sup>12</sup> and the product was obtained after four steps in low yields. Panda and co-workers assembled these compounds via copper [fe](#page-8-0)rrite  $(CuFe<sub>2</sub>O<sub>4</sub>)$ -catalyzed cyclizations.<sup>13</sup> However, these methods either need vigorous reaction conditions or involve multiple steps.

A one-step synthetic route would be a very useful improvement. We have been focusing on the development of a direct synthesis of heterocyclic systems using tandem reactions.<sup>14</sup> Herein, we report an efficient and convergent one-pot synthetic strategy approach to 1,4-thiazepin-5(4H)-one derivativ[es](#page-8-0) under mild conditions. Thiazepine scaffolds were obtained through the reaction of N-substituted 2-mercaptonicotinamides and substituted benzenes at room temperature.

Computational chemistry has been regarded as a valuable tool for elucidating the mechanism of various chemical processes. To improve our understanding of the molecular mechanism of observed Smiles rearrangement products, quantum chemical calculations have been conducted in the framework of density functional theory<sup>15</sup> (DFT) that is one of the most popular tools in electronic structure theory.

## ■ RESULTS AND DISCUSSION

Synthesis. To determine the optimized conditions, the 2 mercapto-N-(p-tolyl)nicotinamide 3c and 1,2-difluoro-4-nitrobenzene 4a were chosen as models. As shown in Table 1, the reaction base, solvent, and time were investigated. The reaction proceeded with different bases in DMSO at room tempe[ra](#page-1-0)ture, and  $Cs_2CO_3$  provided the highest yields (Table 1, entries 1–6). In strongly basic systems such as NaOH or t-KOBu, no desired

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#### <span id="page-1-0"></span>Scheme 1. Structures of Some Biologically Important Thiazepines



Table 1. Optimization of Conditions<sup>a</sup>



a<br>Reaction conditions: 2-mercapto-N-(p-tolyl)nicotinamide 3c (1.0 equiv), 1,2-difluoro-4-nitrobenzene 4a (1.0 equiv), base (3.0 equiv).  $^b$ Isolated yields.

product 5c was obtained (Table 1, entries 1−3). When we used organic base  $Et_3N$  in the reaction, no desired compound  $Sc$  was detected either. To construct  $Sc$ ,  $Cs$ <sub>2</sub>CO<sub>3</sub> performed much better than  $K_2CO_3$  with a yield of 90% (Table 1, entries 5 and 6). The investigation on the solvent proved that the yields of the product in DMSO were higher than in  $CH<sub>3</sub>CN$  with the same base system (Table 1, entries 6 and 7). Finally, we chose  $Cs<sub>2</sub>CO<sub>3</sub>$  in DMSO as the most efficient system to accomplish the synthesis of the 1,4-thiazepin-5(4H)-one derivatives 3 (Table 1, entry 6).

To explore the range of this methodology, various amines were studied (Table 2) under the selected reaction condition (Scheme 2). Structures of products 5a−h are shown in Figure 1. As shown in Table [2](#page-2-0), both aliphatic and aromatic amides led to format[io](#page-2-0)n of tricyclic products in high yields. The reactions [o](#page-3-0)f 1,2-dihalo-4-nitrob[en](#page-2-0)zene 4 with N-aromatic-substituted 2 mercaptonicotinamides (Table 2, entries 5−16) gave yields higher than those of aliphatic 2-mercaptonicotinamides (Table 2, entries 1−4). However, N-ar[om](#page-2-0)atic-substituted amides with electron-donating groups (Table 2, entries 5 and 6) afforded [yi](#page-2-0)elds better than the yields of those with electron-withdrawing groups (Table 2, entries 7 and 8[\).](#page-2-0)

Additionally, the steric hindrance affected the reaction slightly. N-Aromatic-substituted 2-mercapto-N-(3,4,5 trimethoxyphenyl)nicotinamide and N-(3,5-dimethylphenyl)- 2-mercaptonicotinamide (Table 2, entries 13−16) gave high yields of 5 in the reactions. Furthermore, reactions of 2 mercaptonicotinamides with 1,2[-d](#page-2-0)ifluoro-4-nitrobenzene gave the corresponding products in yields higher than the yields of those with 1,2-dichloro-4-nitrobenzene.

As shown in Table 3, a variety of substituted benzenes 4 were used to expand the applicability of this methodology. A substituted benzene-[be](#page-3-0)aring methyl gave a low yield of 40% in the reaction (Table 3, entry 3), whereas benzenes with electron-withdrawing groups produced higher yields (Table 3, entries 1 and 2). The s[tr](#page-3-0)uctures of products 5i−k are shown in Figure 2.

On the basis of our previous work on Smiles rearrangeme[nt](#page-3-0) chemis[try](#page-3-0), a plausible reaction mechanism is presented in Scheme 3. Compounds 3 and 4 underwent nucleophilic substitution, providing compound 8. Then carboxamide anion 9 was for[m](#page-4-0)ed. Subsequently, intermediate 9 could proceed via two pathways (a and b). Pathway b led to 10 by direct intramolecular cyclization. In contrast, pathway a formed intermediate 11 via Smiles rearrangement followed by an

## <span id="page-2-0"></span>Table 2. Synthesis of 9-Nitrobenzo[b]pyrido[3,2-f][1,4]thiazepin-5(6H)-ones<sup>a</sup>



 $^a$ Reaction conditions: 2-mercaptonicotinamides 3 (1.0 equiv), 1,2-dihalo-4-nitrobenzene 4 (1.0 equiv),  $\rm{Cs_2CO_3}$  (3.0 equiv).  $^b$ Isolated yields.  $^c$ Pr and Bu stand for propyl and butyl, respectively.

#### Scheme 2. Synthesis of 1,4-Thiazepin-5(4H)-one Derivatives



intramolecular nucleophilic substitution with a loss of a fluorine atom, leading to the corresponding product 5. To demonstrate this mechanism, the molecular structure of product 5e was determined by X-ray crystallographic analysis (Figure 3).

In addition, to extend the substrate scope, 2-mercaptobenzamides 6 were treated with different substituted [ar](#page-4-0)omatic compounds 4, and the results are shown in Table 4. A variety of N-aryl-substituted 1,4-thiazepin-5(4H)-ones 7 were obtained in good to excellent yields.

Theoretical Results. Scheme 3 schematica[lly](#page-5-0) shows the Smiles rearrangement pathway (pathway a) and the direct nucleophilic substitution pathway ([pa](#page-4-0)thway b). Table 5 gathers optimized geometries of minimal and transition states along two pathways, and Figure 4 gives calculated relative fr[ee](#page-6-0) energy profiles. R denotes the most stable geometry of the anionic reactant.

However, to perform s[ub](#page-7-0)sequent transformation via the ipso-Smiles rearrangement (pathway a) or the direct nucleophilic substitution (pathway b), R must be converted to its conformational isomer, R′, which lies above R by 7.9 kcal/ mol. Intrinsic reaction coordinate (IRC)<sup>5,6</sup> calculations indicate that R′ is the common starting point along with two pathways, in which the negatively charged N3 ato[m i](#page-7-0)s ready to attack C1 or C2.

TS1 corresponds to the transition state in which N3 is nucleophilically attacking the positively charged C1. The forward product from TS1 is IM1, a metastable intermediate

located on the potential energy surface, where the C−N bond has formed and the C1−S bond has broken. The free energy barrier from R to IM1 is calculated to be 14.2 kcal/mol.

 $\mathbf{R}$ 

Once formed, IM1 would be immediately brought to a stable intermediate IM2, which lies below R by 13.7 kcal/mol. Subsequently, IM2 is converted into the precursor of Smiles rearrangement product,  $P_a$ , via TS2 with a free energy barrier of 36.1 kcal/mol. In TS2, the C2−S bond is forming and the C−F bond is breaking. From Figure 4, it is clear that the Smiles rearrangement pathway (pathway a) consists of two elementary steps, and the second step is the [r](#page-7-0)ate-determining step.

Alternatively, if the reaction proceeds via the direct nucleophilic substitution pathway (pathway b), the transition state involved is TS3, which lies above R by 22.6 kcal/mol. Compared to TS1 involved in pathway a, TS3 is energetically less favorable by 8.4 kcal/mol. Therefore, from an energetic point of view, R prefers to evolve along pathway a, resulting in  $P_{a}$ , rather than along pathway b to form  $P_{b}$ . The calculated results are in good agreement with the experimental observation that the ipso-Smiles rearrangement product is substantially predominant over the ortho position nucleophilic substitution product.

## ■ CONCLUSION

In conclusion, a variety of benzo $[b]$ pyrido $[3,2-f][1,4]$ thiazepin-5(6H)-one and dibenzo[ $b_f$ ][1,4]thiazepin-11(10H)-one deriv-

<span id="page-3-0"></span>

Figure 1. Structures of desired compounds 5a−h.





Figure 2. Structures of desired compounds 5i−k.

atives were synthesized in good to excellent yields via Smiles rearrangement. The tricyclic systems were constructed by an efficient one-pot transition metal-free process at room temperature. Furthermore, to gain a better understanding of the reaction mechanism, a theoretical study was performed by quantum chemistry calculations. The results show that the Smiles rearrangement pathway has a barrier that is lower by 14.2 kcal/mol<sup>-1</sup>; thus, it is energetically more favorable than the direct intramolecular nucleophilic substitution pathway. This green and clean synthetic methodology has potential applications in the synthesis of biologically and medicinally relevant compounds.

## **EXPERIMENTAL SECTION**

**General Procedure.** <sup>1</sup>H and <sup>13</sup>CNMR spectra were recorded with a 300 spectrometer or a 400 spectrometer in CDCl<sub>3</sub>. HRMS spectra were recorded on a Q-TOF spectrograph. Compounds 3a−h were prepared according to the literature. Other reagents (Adamas) were commercially available and were used without further purification. All reactions were monitored by thin-layer chromatography (TLC).

Computational Methods. All calculations were conducted using the B3LYP functional<sup>16,17</sup> combined with the 6-311+(d,p) basis set, as implemented in Gaussian 03.<sup>18</sup> Vibrational analyses at the same level of theory were also [con](#page-8-0)ducted to confirm all stationary points as minima (zero imaginary freq[uen](#page-8-0)cies) or first-order saddle points (one imaginary frequency) and to provide free energies at 298.15 K. Solvent effects have also been taken into account by calculating the singlepoint energies of the geometries obtained in the gas phase by using the simple self-consistent reaction field (SCRF) method<sup>19</sup> based on the polarizable continuum (PCM)<sup>20</sup> model. Dimethyl sulfoxide (DMSO)

Table 3. Synthesis of Benzo[b]pyrido[3,2-f][1,4]thiazepin-5(6H)-one Derivatives<sup>a</sup>



a<br>Reaction conditions: 2-mercaptonicotinamides 3 (1.0 equiv), substituted benzenes 4 (1.0 equiv),  $\rm{Cs_2CO_3}$  (3.0 equiv).  $\rm{^{16}Ks_1}$ 

<span id="page-4-0"></span>Scheme 3. ipso-Smiles Rearrangement (pathway a) versus the Direct Nucleophilic Substitution at the Ortho Position (pathway b)





Figure 3. X-ray structure of compound 5e.

was employed as a solvent, corresponding to the experimental conditions.

General Experimental Procedure for 9-Nitro-6-propylbenzo- [b]pyrido[3,2-f][1,4]thiazepin-5(6H)-one (5a). To a solution of DMSO (15 mL) were added 2-mercapto-N-propylnicotinamide 3a (0.20 g, 1.0 mmol), 1,2-difluoro-4-nitrobenzene 4a (0.08 g, 0.5 mmol), and  $Cs_2CO_3$  (0.98 g, 3.0 mmol), and the mixture was stirred for 1 h at room temperature; then another 1,2-difluoro-4-nitrobenzene 4a (0.08 g, 0.5 mmol) was added and the mixture stirred at rt for 4 h. Brine (40 mL) was poured into the solution, and the mixture was extracted with  $CH_2Cl_2$  (3 × 40 mL). The organic layers were combined and dried with anhydrous  $Na<sub>2</sub>SO<sub>4</sub>$ . The product was purified by flash chromatography on silica gel (5:1 hexane/EtOAc). Compound 5a was obtained as a white solid (0.24 g, 0.78 mmol): mp 153–155 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  8.61 (d, J = 2.7 Hz, 1H), 8.50 (dd, J = 4.8, 1.8 Hz, 1H), 8.22 (dd,  $J = 9.0$ , 2.7 Hz, 1H), 8.02 (dd,  $J = 7.8$ , 1.8 Hz, 1H), 7.54 (d, J = 9.0 Hz, 1H), 7.34 (dd, J = 7.5, 4.8 Hz, 1H), 4.65−4.75 (m, 1H), 3.60−3.69 (m, 1H), 1.59−1.78 (m, 2H), 0.96 (t, J  $= 7.4$  Hz, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  166.5, 157.4, 151.3, 148.3, 144.9, 140.1, 135.1, 133.8, 129.5, 126.1, 124.9, 124.0, 53.0, 21.5, 11.4; HRMS calcd for  $C_{15}H_{13}N_3O_3S$   $[(M + H)^+]$  316.0750, found 316.0747.

General Experimental Procedure for 7-Nitro-10-(p-tolyl) dibenzo[b,f][1,4]thiazepin-11(10H)-one (7a). This compound was prepared in the same way as described for 5a by using 2 mercapto-N-(p-tolyl)benzamide (0.24 g, 1.0 mmol), 1,2-difluoro-4 nitrobenzene 4a (0.16 g, 1.0 mmol), and  $Cs_2CO_3$  (0.98 g, 3.0 mmol) in DMSO (15 mL) at room temperature. The product was purified by flash chromatography on silica gel (5:1 hexane/EtOAc) to afford 7a (0.34 g, 0.9 mmol) as a white solid: mp 62–64 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  8.53 (d, J = 2.6 Hz, 1H), 7.97 (dd, J = 9, 2.6 Hz, 1H), 7.79−7.81 (m, 1H), 7.51−7.54 (m, 1H), 7.39−7.42 (m, 2H), 7.24− 7.29 (m, 4H), 7.13 (d, J = 9.0 Hz, 1H), 2.40 (s, 3H); 13C NMR  $(CDCl<sub>3</sub>, 100 MHz)$  δ 168.3, 150.2, 144.2, 140.2, 138.3, 138.1, 137.1, 135.7, 131.8, 131.7, 131.3, 130.4, 129.4, 128.4, 128.1, 124.2, 21.2; FT-HRMS (ESI) calcd for  $C_{20}H_{14}N_2O_3S$   $[(M + H)^+]$  363.0798, found 363.0803.

X-ray Crystal Structure Analysis of Compound 5e.<sup>21</sup> Single crystals of 5e suitable for X-ray crystal analysis were obtained by recrystallization from a hexane/ $CH_2Cl_2$  mixed solvent. Inte[nsi](#page-8-0)ty data were collected at 293 K on an X-ray diffractometer with Mo Kα radiation ( $\lambda = 0.71073$  Å) and a graphite monochromator. A total of 9276 reflections were measured at a maximal  $2\theta$  angle of  $50.0^{\circ}$ , of which 3792 were independent reflections  $(R<sub>int</sub> = 0.0500)$ . The structure was determined by direct methods  $(SHELXS-97)^{22}$  and refined by the full-matrix least-squares method on  $F^2$  (SHELEXL-97).<sup>22</sup> The crystal data are as follows:  $C_{18}H_{10}CIN_3O_3S$ ,  $F_W = 383.01$  $F_W = 383.01$  $F_W = 383.01$ , crystal size of 0.12 mm  $\times$  0.10 mm  $\times$  0.10 mm, monoclinic, P121/c1, a  $= 8.784(3)$  $= 8.784(3)$  $= 8.784(3)$  Å,  $b = 16.217(5)$  Å,  $c = 14.313(4)$  Å,  $V = 1700.5(9)$  Å<sup>3</sup>,  $Z =$ 4. The refinement converged to  $R_1 = 0.0467$ ,  $wR_2 = 0.1352$   $[I > 2\sigma(I)],$ and GOF = 1.111.

6-Butyl-9-nitrobenzo[b]pyrido[3,2-f ][1,4]thiazepin-5(6H) **one (5b):** 264 mg (80% yield); pale yellow oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  8.60 (d,  $J = 1.8$  Hz, 1H), 8.49 (dd,  $J = 4.8$ , 1.2 Hz, 1H), 8.22  $(dd, J = 8.7, 2.1 Hz, 1H), 8.01 (dd, J = 7.8, 1.2 Hz, 1H), 7.55 (d, J =$ 9.0 Hz, 1H), 7.33 (d, J = 7.8, 5.1 Hz, 1H), 4.69−4.79 (m, 1H), 3.62− 3.71 (m, 1H), 1.58−1.69 (m, 2H), 1.32−1.1.44 (m, 2H), 0.90 (t, J = 7.2 Hz, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 166.4, 157.4, 151.3, 148.3, 144.9, 140.1, 135.1, 133.8, 129.4, 126.1, 124.9, 124.0, 51.0, 30.2, 20.1, 13.7; FT-HRMS (ESI) calcd for  $C_{16}H_{15}N_3O_3S$   $[(M + H)^+]$  330.0907, found 330.0933.

9-Nitro-6-(p-tolyl)benzo[b]pyrido[3,2-f ][1,4]thiazepin-5(6H) one (5c): 326 mg (90% yield); white solid; mp 185−187 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  8.64 (d, J = 2.7 Hz, 1H), 8.55 (dd, J = 4.8, 1.8 Hz, 1H), 8.12 (dd, J = 7.8, 1.8 Hz, 1H), 8.02 (dd, J = 9.0, 2.7 Hz, 1H), 7.39 (dd, J = 7.8, 4.8 Hz, 1H), 7.24−7.32 (m, 5H), 7.19 (d, J = 9.0 Hz, 1H), 2.41 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  166.5, 157.0, 151.7, 149.1, 144.6, 140.2, 139.5, 138.7, 134.1, 133.2, 130.4, 129.4, 128.1, 124.5, 124.1, 21.2; FT-HRMS (ESI) calcd for  $C_{19}H_{13}N_3O_3S$  [(M + H)<sup>+</sup>] 364.0750, found 364.0766.

6-(4-Fluorophenyl)-9-nitrobenzo[b]pyrido[3,2-f ][1,4] **thiazepin-5(6H)-one (5d):** 322 mg (88% yield); white solid; mp 148−150 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  8.65 (d, J = 2.4 Hz, 1H),

## <span id="page-5-0"></span>Table 4. Synthesis of 1,4-Thiazepin-5(4H)-one Derivatives<sup>a</sup>



a<br>Reaction conditions: 2-mercaptonicotinamides 3 (1.0 equiv), substituted benzenes 4 (1.0 equiv),  $\rm{Cs_2CO_3}$  (3.0 equiv).  $^b$ Isolated yields.

8.56 (d,  $J = 3.3$  Hz, 1H), 8.13 (d,  $J = 1.2$  Hz, 1H), 8.05 (dd,  $J = 9.0, 2.7$ Hz, 1H), 7.36–7.42 (m, 3H), 7.15–7.22 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  166.5, 162.0 ( $^{1}J_{C,F}$  = 248 Hz), 157.0, 151.9, 148.8, 144.8, 140.3, 137.9  $({}^{4}J_{C,F} = 3 \text{ Hz})$ , 133.5  $({}^{2}J_{C,F} = 19 \text{ Hz})$ , 130.2  $({}^{3}J_{C,F} = 9 \text{ Hz})$ , 129.5, 128.0, 124.7, 124.2, 117.0, 116.6; FT-HRMS (ESI) calcd for  $C_{18}H_{10}FN_3O_3S$  [(M + H)<sup>+</sup>] 368.0500, found 368.0516.

6-(4-Chlorophenyl)-9-nitrobenzo[b]pyrido[3,2-f ][1,4] thiazepin-5(6H)-one (5e): 363 mg (95% yield); white solid; mp 195−196 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.65 (d, J = 2.7 Hz, 1H), 8.56 (dd, J = 4.8, 1.8 Hz, 1H), 8.11 (dd, J = 7.8, 1.8 Hz, 1H), 8.05 (dd, J = 9, 2.7 Hz, 1H), 7.45−7.50 (m, 2H), 7.39 (dd, J = 7.8, 4.8 Hz, 1H), 7.31−7.36 (m, 2H), 7.16 (d, J = 9.0 Hz, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 166.3, 157.0, 151.9, 148.6, 144.9, 140.4, 140.3, 134.4, 133.7,

<span id="page-6-0"></span>Table 5. Optimized Structures for Transition States, Intermediates, and Products Involved along Pathways a and b Shown in Scheme  $3^a$ 



a Distances are in angstroms.

133.6, 130.0, 129.7, 129.5, 128.1, 124.7, 124.2; FT-HRMS (ESI) calcd for  $C_{18}H_{10}CIN_3O_3S$   $[(M + H)^+]$  384.0204, found 384.0214.

6-(4-Bromophenyl)-9-nitrobenzo[b]pyrido[3,2-f ][1,4] thiazepin-5(6H)-one (5f): 388 mg (91% yield); white solid; mp 216−218 °C; <sup>1</sup> H NMR (CDCl3, 300 MHz) δ 8.65 (d, J = 2.7 Hz, 1H), 8.56 (dd, J = 4.8, 1.8 Hz, 1H), 8.11 (dd, J = 7.8, 1.8 Hz, 1H), 8.05 (dd, J = 9.0, 2.7 Hz, 1H), 7.60−7.65 (m, 2H), 7.39 (dd, J = 7.8, 4.8 Hz, 1H), 7.25−7.29 (m, 2H), 7.15 (d, J = 9.0 Hz, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 166.2, 157.0, 151.9, 148.5, 144.9, 141.0, 140.3, 133.7, 133.0, 130.0, 129.5, 128.1, 124.7, 124.2, 122.5; FT-HRMS (ESI) calcd for  $C_{18}H_{10}BrN_3O_3S$   $[(M + H)^+]$  427.9699, found 427.9673.

6-(3,5-Dimethylphenyl)-9-nitrobenzo[b]pyrido[3,2-f ][1,4] thiazepin-5(6H)-one (5g): 358 mg (95% yield); white solid; mp none; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  8.64 (d, J = 2.4 Hz, 1H), 8.55  $(d, J = 1.4 \text{ Hz}, 1H), 8.10 (d, J = 7.7 \text{ Hz}, 1H), 8.02 (dd, J = 9.0, 2.4 \text{ Hz},$ 1H), 7.38 (dd, J = 7.7, 2.9 Hz, 1H), 7.22 (d, J = 9.0 Hz, 1H), 7.04 (s, 1H), 6.99 (s, 2H), 2.36 (s, 6H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  166.4, 157.0, 151.6, 149.0, 144.6, 141.9, 140.1, 139.7, 134.2, 133.1, 130.4, 129.4, 128.0, 126.0, 124.5, 124.1, 21.2; FT-HRMS (ESI) calcd for  $C_{20}H_{15}N_3O_3S$  [(M + H)<sup>+</sup>] 378.0907, found 378.0919.

9-Nitro-6-(3,4,5-trimethoxyphenyl)benzo[b]pyrido[3,2-f ]- [1,4]thiazepin-5(6H)-one (5h):  $395 \text{ mg}$  (90% yield); white solid; mp 228−230 °C; <sup>1</sup> H NMR (CDCl3, 300 MHz) δ 8.65 (d, J = 2.7 Hz, 1H), 8.56 (dd, J = 4.8, 1.8 Hz, 1H), 8.12 (dd, J = 7.8, 1.8 Hz, 1H), 8.07 (dd,  $J = 9.0, 2.7 \text{ Hz}, 1\text{H}$ , 7.39 (dd,  $J = 7.8, 4.8 \text{ Hz}, 1\text{H}$ ), 7.30 (d,  $J = 9.0 \text{ Hz}$ , 1H), 6.58 (s, 2H), 3.89 (s, 3H), 3.87 (s, 6H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 166.5, 156.9, 154.0, 151.8, 148.8, 144.7, 140.2, 138.3, 137.4, 134.0, 132.9, 129.5, 127.7, 124.7, 124.2, 106.0, 60.9, 56.4; FT-HRMS (ESI) calcd for  $C_{21}H_{17}N_3O_6S$  [(M + H)<sup>+</sup>] 440.0872, found 440.0853.

9-Fluoro-6-(p-tolyl)benzo[b]pyrido[3,2-f ][1,4]thiazepin-5(6H)-one (5i): 278 mg (83% yield); white solid; mp 152−153 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  8.51 (dd, J = 4.8, 1.8 Hz, 1H), 8.12  $(dd, J = 7.8, 1.8 Hz, 1H), 7.46 (dd, J = 7.8, 3 Hz, 1H), 7.35 (dd, J =$ 7.8, 4.8 Hz, 1H), 7.26 (s, 5H), 7.06 (dd, J = 9.0, 5.1 Hz, 1H), 6.87−

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Figure 4. Calculated relative free energy profiles along the ipso-Smiles rearrangement pathway (pathway a) and the direct nucleophilic substitution pathway (pathway b).

6.94 (m, 1H), 2.39 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  172.3, 166.7, 162.8  $({}^{1}J_{C,F} = 232 \text{ Hz})$ , 158.1, 157.8, 156.1, 150.9, 140.1  $({}^{3}J_{C,F} =$ 9 Hz), 140.0, 137.8, 134.5, 130.1, 129.1 ( ${}^{3}J_{C,F}$  = 8 Hz), 127.8, 123.8, 120.4 ( ${}^{2}J_{\text{C,F}}$  = 22 Hz), 117.1 ( ${}^{2}J_{\text{C,F}}$  = 22 Hz), 21.2; <sup>19</sup>F NMR (CDCl<sub>3</sub>, 282 MHz)  $\delta$  –114.23; FT-HRMS (ESI) calcd for C<sub>19</sub>H<sub>13</sub>FN<sub>2</sub>OS [(M + H)+ ] 337.0766, found 337.0790.

6-(3,5-Dimethylphenyl)-9-fluorobenzo[b]pyrido[3,2-f ][1,4] thiazepin-5(6H)-one (5j): 315 mg (93% yield); white solid; mp 148−150 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.52 (dd, J = 4.8, 1.5 Hz, 1H), 8.12 (d, J = 4.8 Hz, 1H), 7.46 (dd, J = 7.8, 2.7 Hz, 1H), 7.36 (dd, J = 7.8, 4.8 Hz, 1H), 7.10 (dd, J = 9.0, 5.1 Hz, 1H), 7.00 (s, 3H), 6.88− 6.95 (m, 1H), 2.35 (s, 6H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  165.6, 157.0,156.8, 149.8, 141.5, 139.2, 138.9  $({}^{4}J_{C,F} = 4$  Hz), 138.3, 133.6, 133.1 ( ${}^{3}J_{C,F}$  = 9 Hz), 128.7, 128.1 ( ${}^{3}J_{C,F}$  = 9 Hz), 124.7, 122.9, 119.5  $(^{2}J_{\text{C,F}} = 23 \text{ Hz})$ , 116.1  $(^{2}J_{\text{C,F}} = 22 \text{ Hz})$ , 20.3; <sup>19</sup>F NMR (CDCl<sub>3</sub>, 282) MHz)  $\delta$  -114.36; FT-HRMS (ESI) calcd for C<sub>20</sub>H<sub>15</sub>FN<sub>2</sub>OS [(M + H)+ ] 351.0962, found 351.0972.

6-(3,5-Dimethylphenyl)-7-methylbenzo[b]pyrido[3,2-f ][1,4] **thiazepin-5(6H)-one (5k):** 138 mg  $(40\% \text{ yield})$ ; white solid; mp 190−192 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.43 (dd, J = 4.8, 1.8 Hz, 1H), 8.06 (dd, J = 7.5, 1.8 Hz, 1H), 7.68 (dd, J = 7.2, 2.4 Hz, 1H), 7.30 (dd, J = 7.8, 4.8 Hz, 1H), 7.08–7.15 (m, 2H), 6.98 (s, 2H), 6.86 (s, 1H), 2.30 (s, 6H), 2.00 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ 166.8, 150.0, 141.4, 140.8, 139.5, 137.9, 137.3, 136.8, 135.1, 132.9, 132.1, 128.2, 127.4, 124.7, 123.6, 21.4, 20.0; FT-HRMS (ESI) calcd for  $C_{21}H_{18}N_2OS [(M + H)^+]$  347.1213, found 347.1224.

9-(Tetrahydro-2H-pyran-2-yl)-6-(p-tolyl)pyridazino[4,5-b] pyrido[3,2-f ][1,4]thiazepine-5,10(6H,9H)-dione (7b): 398 mg (95% yield); white crystal; mp 98-100 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 7.77 (m, 1H), 7.52−7.57 (m, 1H), 7.39−7.46 (m, 2H), 7.35 (s, 1H), 7.22−7.31 (m, 4H), 5.99 (dd, J = 10.5, 1.8 Hz, 1H), 4.07 (d, J = 11.4 Hz, 1H), 3.68−3.76 (m, 1H), 2.40 (s, 3H), 1.92−2.09 (m, 2H), 1.51−1.76 (m, 4H); 13C NMR (CDCl3, 75 MHz) δ 168.7, 145.3, 139.2, 138.1, 137.0, 136.9, 134.4, 132.4, 132.0, 131.9, 130.5, 129.3, 128.3, 83.6, 68.8, 28.6, 24.8, 22.7, 21.2; FT-HRMS (ESI) calcd for  $C_{23}H_{21}N_3O_3S$  [(M + H)<sup>+</sup>] 420.1376, found 420.1382.

10-(4-Chlorophenyl)-7-nitrodibenzo[b,f ][1,4]thiazepin-11- (10H)-one (7c): 324 mg (85% yield); white solid; mp 98–100 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  8.54 (d, J = 2.6 Hz, 1H), 8.00 (dd, J = 8.9, 2.6 Hz, 1H), 7.78−7.81 (m, 1H), 7.53−7.55 (m, 1H), 7.40−7.46 (m, 4H), 7.33 (d, J = 8.7 Hz, 1H), 7.10 (d, J = 9.0 Hz, 1H); 13C NMR (CDCl3, 100 MHz) δ 168.1, 149.7, 144.5, 141.1, 137.6, 137.1, 136.2, 134.1, 131.9, 131.8, 131.4, 130.0, 129.9, 129.5, 128.6, 128.1, 124.4; FT-HRMS (ESI) calcd for  $C_{19}H_{11}CIN_2O_3S$   $[(M + H)^+]$  383.0252, found 383.0253.

11-(4-Chlorophenyl)-3-(tetrahydro-2H-pyran-2-yl)benzo[f ] pyridazino[4,5-b][1,4]thiazepine-4,10(3H,11H)-dione (7d): 390 mg (89% yield); white crystal; mp 138–140 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.79 (d, J = 4.9 Hz, 1H), 7.55 (d, J = 3.7 Hz, 1H), 7.43–

7.52 (m, 4H), 5.98 (d,  $J = 10.2$  Hz, 1H), 4.07 (d,  $J = 11.4$  Hz, 1H), 7.31 (d, J = 4.0 Hz, 3H) 1.98–2.05 (m, 2H), 1.52–1.70 (m, 4H); <sup>13</sup>C NMR (CDCl3, 100 MHz) δ 168.5, 158.4, 144.9, 138.1, 137.7, 137.0, 135.0, 134.2, 132.6, 132.3, 132.1, 130.1, 130.0, 129.4. 83.8, 68.9, 28.7, 24.8, 22.7; FT-HRMS (ESI) calcd for  $C_{22}H_{18}CIN_3O_3S$   $[(M + H)^+]$ 440.0830, found 440.0864.

## ■ ASSOCIATED CONTENT

#### **8** Supporting Information

Copies of  ${}^{1}H$  and  ${}^{13}C$  NMR spectra of all compounds, X-ray data of 5e in CIF format, and Cartesian coordinates and absolute energies for all structures involved in theoretical calculations. This material is available free of charge via the Internet at http://pubs.acs.org.

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#### Notes

The auth[ors declare no comp](mailto:zhangdj@sdu.edu.cn)eting financial interest.

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#### ■ REFERENCES

(1) (a) Masumiya, H.; Tanaka, H.; Shigenobu, K. Eur. J. Pharmacol. 1997, 335, 15−21. (b) Watanabe, T.; Kalasz, H.; Yabana, H.; Kuniyasu, A.; Mershon, J.; Itagaki, K.; Vaghy, P. L.; Naito, K.; Nakayama, H.; Schwartz, A. FEBS Lett. 1993, 334, 261−264. (c) Bariwal, J. B.; Upadhyay, K. D.; Manvar, A. T.; Trivedi, J. C.; Singh, J. S.; Jain, K. S.; Shah, A. K. Eur. J. Med. Chem. 2008, 43, 2279− 2290. (d) Wajima, T.; Fukumura, K.; Yano, Y.; Oguma, T. J. Pharm. Sci. 2003, 92, 2427−2440. (e) Beaucage, P.; Massicotte, J.; Boileau, J. F.; Dumont, L. J. Cardiovasc. Pharmacol. 2003, 42, 142−150.

(2) (a) Geller, V.; Gorzaltsan, I.; Shleifer, T.; Belmaker, R. H.; Bersudsky, Y. Schizophr. Res. Treat. 2005, 80, 343−347. (b) Kurita, M.; Holloway, T.; García-Bea, A.; Kozlenkov, A.; Friedman, A. K.; Moreno, J. L.; Heshmati, M.; Golden, S. A.; Kennedy, P. J.; Takahashi, N.; Dietz, D. M.; Mocci, G.; Gabilondo, A. M.; Hanks, J.; Umali, A.; Callado, L. F.; Gallitano, A. L.; Neve, R. L.; Shen, L.; Buxbaum, J. D.; Han, M. H.; Nestler, E. J.; Meana, J. J.; Russo, S. J.; González-Maeso, J. Nat. Neurosci. 2012, 15, 1245−1254. (c) Kapur, S.; Seeman, P. Am. J. Psychiatry 2001, 158, 360. (d) Richelson, E.; Souder, T. Life Sci. 2000, 68, 29−39. (e) Seminara, G.; Trassari, V.; Prestifilippo, N.; Chiavetta, R.; Calandra, C. Minerva Psichiatrica 1993, 34, 95−99.

(3) Pettersson, H.; Bülow, A.; Ek, F.; Jensen, F. E.; Otteson, L. K.; Fejzic, A.; Ma, J.-N.; Del Tredici, A. L.; Currier, E. A.; Gardell, L. R.; Tabatabaei, A.; Craig, D.; McFarland, K.; Ott, T. R.; Piu, F.; Burstein, E. S.; Olsson, R. J. Med. Chem. 2009, 52, 1975−1982.

(4) Binaschi, M.; Boldetti, A.; Gianni, M.; Maggi, C. A.; Gensini, M.; Bigioni, M.; Parlani, M.; Giolitti, A.; Fratelli, M.; Valli, C.; Terao, M.; Garattini, E. ACS Med. Chem. Lett. 2010, 1, 411−415.

(5) (a) Kumar, M.; Sharma, K.; Fogla, A. K.; Sharma, K.; Rathore, M. Res. Chem. Intermed. 2013, 39, 2555−2564. (b) Anisetti, R.; Reddy, M. S. J. Sulfur Chem. 2012, 33, 363−372. (c) Sayed, G. H.; Sayed, M. A.; Mahmoud, M. R.; Shaaban, S. S. Egypt. J. Chem. 2002, 45, 767−776. (d) Patel, R. N.; Nimavat, K. S.; Vyas, K. B.; Patel, P. V. J. Chem. Pharm. Res. 2011, 3, 409−415.

(6) Frolov, E. B.; Lakner, F. J.; Khvat, A. V.; Ivachtchenko, A. V. Tetrahedron Lett. 2004, 45, 4693−4696.

<span id="page-8-0"></span>(7) (a) Robl, J. A.; Sun, C. Q.; Stevenson, J.; Ryono, D. E.; Simpkins, L. M.; Cimarusti, M. P.; Dejneka, T. W.; Slusarchyk, A.; Chao, S.; Stratton, L.; Misra, R. N.; Bednarz, M. S.; Asaad, M. M.; Cheung, H. S.; Abboa-Offei, B. E.; Smith, P. L.; Mathers, P. D.; Fox, M.; Schaeffer, T. R.; Seymour, A. A.; Trippodo, N. C. J. Med. Chem. 1997, 40, 1570− 1577. (b) Barbaro, R.; Betti, L.; Botta, M.; Corelli, F.; Giannaccini, G.; Maccari, L.; Manetti, F.; Strappaghetti, G.; Corsano, S. J. Med. Chem. 2001, 44, 2118−2132. (c) Oda, T.; Tijima, Y.; Sada, T.; Nishino, H.; Oizumi, K.; Koike, H. Biochem. Biophys. Res. Commun. 1988, 152, 456−462.

(8) Katrusiak, B.; Unlu, S.; Banoglu, E.; Kupeli, E.; Yesilada, E.; Sahin, M. F. Arch. Pharm. Pharm. Med. Chem. 2003, 336, 406−412.

(9) Libgeois, J. F.; Rogistert, F. A.; Bruhwyler, J.; Damas, J.; Nguyen,

T. P.; Inarejos, M. O.; Eric, M. G. J. Med. Chem. 1994, 37, 519−525. (10) Fu, R.; Xiu, X.; Dang, Q.; Bai, X. J. Org. Chem. 2005, 70, 10810− 10816.

(11) Binaschi, M.; Boldetti, A.; Gianni, M.; Maggi, C. A.; Gensini, M.; Bigioni, M.; Parlani, M.; Giolitti, A.; Fratelli, M.; Valli, C.; Terao, M.; Garattini, E. ACS Med. Chem. Lett. 2010, 1, 411−415.

(12) Ottesen, L. K.; Fredrik, E.; Roger, O. Org. Lett. 2006, 8, 1771− 1773.

(13) Panda, N.; Jena, A. K.; Mohapatra, S. Appl. Catal., A 2012, 433− 434, 258−264.

(14) (a) Huang, A. P.; Chen, Y. M.; Zhou, Y. G.; Guo, W.; Wu, X. D.; Ma, C. Org. Lett. 2013, 15, 5480−5483. (b) Zhao, Y. M.; Wu, Y. M.; Jia, J.; Zhang, D. J.; Ma, C. J. Org. Chem. 2012, 77, 8501−8506. (c) Yang, B. C.; Niu, X. Y.; Huang, Z. X.; Zhao, C. H.; Liu, Y.; Ma, C. Tetrahedron 2013, 69, 8250−8254. (d) Yang, B. C.; Huang, Z. X.; Guan, H. G.; Niu, X. Y.; Li, Y. Q.; Fang, S.; Ma, C. Tetrahedron Lett. 2013, 54, 5994−5997. (e) Niu, X. Y.; Yang, B. C.; Li, Y. Q.; Fang, S.; Huang, Z. X.; Xie, C. X.; Ma, C. Org. Biomol. Chem. 2013, 11, 4102− 4108. (f) Liu, Y. L.; Zhan, C. J.; Yang, B. C.; Cao, X. Q.; Ma, C. Synlett 2013, 45, 111−117. (g) Liu, Y. L.; Chu, C. X.; Huang, A. P.; Zhan, C. J.; Ma, Y.; Ma, C. ACS Comb. Sci. 2011, 13, 547−553.

(15) Kohn, W.; Becke, A. D.; Parr, R. G. J. Phys. Chem. 1996, 100, 12974−12980.

(16) Lee, C.; Yang, W.; Parr, G. Phys. Rev. B 1988, 37, 785−794.

(17) Becke, A. D. J. Chem. Phys. 1993, 98, 5648−5652.

(18) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Montgomery, J. A., Jr.; Vreven, T.; Kudin, K. N.; Burant, J. C.; Millam, J. M.; Iyengar, S. S.; Tomasi, J.; Barone, V.; Mennucci, B.; Cossi, M.; Scalmani, G.; Rega, N.; Petersson, G. A.; Nakatsuji, H.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Klene, M.; Li, X.; Knox, J. E.; Hratchian, H. P.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Ayala, P. Y.; Morokuma, K.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Zakrzewski, V. G.; Dapprich, S.; Daniels, A. D.; Strain, M. C.; Farkas, O.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Ortiz, J. V.; Cui, Q.; Baboul, A. G.; Clifford, S.; Cioslowski, J.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen, W.; Wong, M. W.; Gonzalez, C.; Pople, J. A. Gaussian 03; Gaussian, Inc.: Wallingford, CT, 2004.

(19) Tapia, O. J. Math. Chem. 1992, 10, 139−181.

(20) Barone, V.; Cossi, M.; Tomasi, J. J. Comput. Chem. 1998, 19, 404−407.

(21) For the crystal data of 5e, see the Supporting Information. CCDC 994893 (5e) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crysallographic Data Centre [via www.ccdc.cam.ac.uk/](#page-7-0) data\_request/cif.

(22) Sheldrick, G. M. SHELX-97, Program for the Refinement of [Crystal Structure](www.ccdc.cam.ac.uk/data_request/cif); University of Göttingen: Götti[ngen, Germany, 1997.](www.ccdc.cam.ac.uk/data_request/cif)