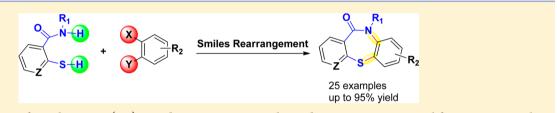
Transition Metal-Free One-Pot Synthesis of Fused 1,4-Thiazepin-5(4*H*)-ones and Theoretical Study of the S–N Type Smiles Rearrangement Process

Bingchuan Yang,[†] Xiaochen Tan,[†] Ruiying Guo,[†] Shunwei Chen,[†] Zeyuan Zhang,[†] Xianglong Chu,[†] Caixia Xie,[†] Dongju Zhang,^{*,†} and Chen Ma^{*,†,‡}

[†]School of Chemistry and Chemical Engineering, Shandong University, Jinan 250100, P. R. China

[‡]State Key Laboratory of Bioactive Substance and Function of Natural Medicines, Institute of Materia Medica, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing 100050, P. R. China

Supporting Information



ABSTRACT: A series of 1,4-thiazepin-5(4*H*)-one derivatives were synthesized via a transition metal-free one-pot Smiles rearrangement process at room temperature. Regioselective seven-membered heterocycles were constructed in good to excellent yields. To gain an in-depth understanding of the S-N type Smiles rearrangement mechanism, a theoretical study was also performed by quantum chemistry calculations.

INTRODUCTION

1,4-Thiazepin-5(4*H*)-one derivatives have attracted a great deal of interest because of their outstanding biological and medicinal properties. Compounds containing thiazepine functionalities have been broadly used to cure all kinds of diseases. For example, clentiazem is an important calcium channel blocker, used in the treatment of angina pectoris and hypertension.¹ Clotiapine acts as an antipsychotic for the treatment of bipolar disorder, depressive disorder, and schizophrenia.² Ethyl 4-(11-propyldibenzo[b_if][1,4]thiazepine-2-carboxamido)piperidine-1-carboxylate (1) was reported as a potential inverse agonist.³ *N*-Hydroxy-6-(11-oxodibenzo[b_if][1,4] thiazepine-10(11*H*)-yl)-hexanamide (**2**) and its analogues exhibit high antitumor activities (Scheme 1).⁴ Fused thiazepine scaffolds also have antimicrobial,⁵ anti-inflammatory,⁶ antihypertensive,⁷ and analgesic activities.⁸

As a result, various methods have been explored for the synthesis of 1,4-thiazepin-5(4*H*)-one scaffolds, including the Friedel–Crafts reaction beginning with 4-chlorobenzenethiol and 2-chloronicotinic acid,⁹ the heating of 4,6-dichloropyr-imidin-5-amine with benzenethiols,¹⁰ and the copper-catalyzed Ullmann arylations using 2-mercaptobenzoic acid and 1-bromo-4-methoxy-2-nitrobenzene.¹¹ Ottesen also reported the synthesis of 1,4-thiazepin-5(4*H*)-one derivatives by coupling 4-chloro-1-fluoro-2-nitrobenzene with 2-mercaptobenzoate,¹² and the product was obtained after four steps in low yields. Panda and co-workers assembled these compounds via copper ferrite (CuFe₂O₄)-catalyzed cyclizations.¹³ However, these methods

either need vigorous reaction conditions or involve multiple steps.

A one-step synthetic route would be a very useful improvement. We have been focusing on the development of a direct synthesis of heterocyclic systems using tandem reactions.¹⁴ Herein, we report an efficient and convergent one-pot synthetic strategy approach to 1,4-thiazepin-5(4H)-one derivatives under mild conditions. Thiazepine scaffolds were obtained through the reaction of N-substituted 2-mercaptonicotinamides and substituted benzenes at room temperature.

Computational chemistry has been regarded as a valuable tool for elucidating the mechanism of various chemical processes. To improve our understanding of the molecular mechanism of observed Smiles rearrangement products, quantum chemical calculations have been conducted in the framework of density functional theory¹⁵ (DFT) that is one of the most popular tools in electronic structure theory.

RESULTS AND DISCUSSION

Synthesis. To determine the optimized conditions, the 2mercapto-N-(p-tolyl)nicotinamide 3c and 1,2-difluoro-4-nitrobenzene 4a were chosen as models. As shown in Table 1, the reaction base, solvent, and time were investigated. The reaction proceeded with different bases in DMSO at room temperature, and Cs₂CO₃ provided the highest yields (Table 1, entries 1–6). In strongly basic systems such as NaOH or *t*-KOBu, no desired

 Received:
 May 26, 2014

 Published:
 August 7, 2014

Article

Scheme 1. Structures of Some Biologically Important Thiazepines

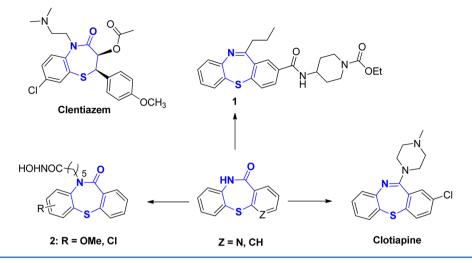


Table 1. Optimization of Conditions^a

	O N SH 3c	$\frac{F}{F} = \frac{Cs_2CO_3, DM}{NO_2}$	$SO \rightarrow N \rightarrow NO_2$	
entry	base	solvent	time (h)	yield (%) ^b
1	КОН	DMSO	8	nd
2	NaOH	DMSO	8	nd
3	t-BuOK	DMSO	8	nd
4	Et ₃ N	DMSO	8	nd
5	K ₂ CO ₃	DMSO	8	76
6	Cs_2CO_3	DMSO	6	90
7	Cs ₂ CO ₃	CH ₃ CN	10	40

^{*a*}Reaction conditions: 2-mercapto-*N*-(*p*-tolyl)nicotinamide 3c (1.0 equiv), 1,2-difluoro-4-nitrobenzene 4a (1.0 equiv), base (3.0 equiv). ^{*b*}Isolated yields.

product **5c** was obtained (Table 1, entries 1–3). When we used organic base Et_3N in the reaction, no desired compound **5c** was detected either. To construct **5c**, Cs_2CO_3 performed much better than K_2CO_3 with a yield of 90% (Table 1, entries 5 and 6). The investigation on the solvent proved that the yields of the product in DMSO were higher than in CH₃CN with the same base system (Table 1, entries 6 and 7). Finally, we chose Cs_2CO_3 in DMSO as the most efficient system to accomplish the synthesis of the 1,4-thiazepin-5(4*H*)-one derivatives **3** (Table 1, entry 6).

To explore the range of this methodology, various amines were studied (Table 2) under the selected reaction condition (Scheme 2). Structures of products 5a-h are shown in Figure 1. As shown in Table 2, both aliphatic and aromatic amides led to formation of tricyclic products in high yields. The reactions of 1,2-dihalo-4-nitrobenzene 4 with N-aromatic-substituted 2mercaptonicotinamides (Table 2, entries 5–16) gave yields higher than those of aliphatic 2-mercaptonicotinamides (Table 2, entries 1–4). However, N-aromatic-substituted amides with electron-donating groups (Table 2, entries 5 and 6) afforded yields better than the yields of those with electron-withdrawing groups (Table 2, entries 7 and 8). Additionally, the steric hindrance affected the reaction slightly. *N*-Aromatic-substituted 2-mercapto-N-(3,4,5-trimethoxyphenyl)nicotinamide and *N*-(3,5-dimethylphenyl)-2-mercaptonicotinamide (Table 2, entries 13–16) gave high yields of **5** in the reactions. Furthermore, reactions of 2-mercaptonicotinamides with 1,2-difluoro-4-nitrobenzene gave the corresponding products in yields higher than the yields of those with 1,2-dichloro-4-nitrobenzene.

As shown in Table 3, a variety of substituted benzenes 4 were used to expand the applicability of this methodology. A substituted benzene-bearing methyl gave a low yield of 40% in the reaction (Table 3, entry 3), whereas benzenes with electron-withdrawing groups produced higher yields (Table 3, entries 1 and 2). The structures of products 5i-k are shown in Figure 2.

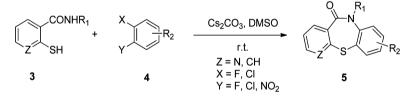
On the basis of our previous work on Smiles rearrangement chemistry, a plausible reaction mechanism is presented in Scheme 3. Compounds 3 and 4 underwent nucleophilic substitution, providing compound 8. Then carboxamide anion 9 was formed. Subsequently, intermediate 9 could proceed via two pathways (a and b). Pathway b led to 10 by direct intramolecular cyclization. In contrast, pathway a formed intermediate 11 via Smiles rearrangement followed by an

Table 2. Synthesis of 9-Nitrobenzo[b]pyrido[3,2-f][1,4]thiazepin-5(6H)-ones^a

	CONHR ₁ +	+ +					
	`N ́``SH	x NO2	r.t.	S NO ₂			
	3	4		5			
entry	\mathbb{R}^1	Х	time (h)	product	yield $(\%)^b$		
1	Pr ^c	F	5	5a	78		
2	Pr ^c	Cl	6	5a	65		
3	Bu ^c	F	5	5b	80		
4	Bu ^c	Cl	6	5b	78		
5	$4-MeC_6H_4$	F	6	5c	90		
6	$4-MeC_6H_4$	Cl	6	5c	73		
7	$4-FC_6H_4$	F	6	5d	88		
8	$4-FC_6H_4$	Cl	6	5d	75		
9	4-ClC ₆ H ₄	F	6	5e	95		
10	4-ClC ₆ H ₄	Cl	6	5e	78		
11	4-BrC ₆ H ₄	F	6	5f	91		
12	4-BrC ₆ H ₄	Cl	6	5f	76		
13	3,5-dimethylphenyl	F	6	5g	95		
14	3,5-dimethylphenyl	Cl	6	5g	80		
15	$3,4,5-(MeO)_3C_6H_2$	F	6	5h	90		
16	$3,4,5-(MeO)_3C_6H_2$	Cl	6	5h	76		

"Reaction conditions: 2-mercaptonicotinamides 3 (1.0 equiv), 1,2-dihalo-4-nitrobenzene 4 (1.0 equiv), Cs₂CO₃ (3.0 equiv). ^bIsolated yields. ^cPr and Bu stand for propyl and butyl, respectively.

Scheme 2. Synthesis of 1,4-Thiazepin-5(4H)-one Derivatives



intramolecular nucleophilic substitution with a loss of a fluorine atom, leading to the corresponding product **5**. To demonstrate this mechanism, the molecular structure of product **5e** was determined by X-ray crystallographic analysis (Figure 3).

In addition, to extend the substrate scope, 2-mercaptobenzamides **6** were treated with different substituted aromatic compounds **4**, and the results are shown in Table 4. A variety of *N*-aryl-substituted 1,4-thiazepin-5(4H)-ones 7 were obtained in good to excellent yields.

Theoretical Results. Scheme 3 schematically shows the Smiles rearrangement pathway (pathway a) and the direct nucleophilic substitution pathway (pathway b). Table 5 gathers optimized geometries of minimal and transition states along two pathways, and Figure 4 gives calculated relative free energy profiles. R denotes the most stable geometry of the anionic reactant.

However, to perform subsequent transformation via the *ipso*-Smiles rearrangement (pathway a) or the direct nucleophilic substitution (pathway b), R must be converted to its conformational isomer, R', which lies above R by 7.9 kcal/ mol. Intrinsic reaction coordinate (IRC)^{5,6} calculations indicate that R' is the common starting point along with two pathways, in which the negatively charged N3 atom is ready to attack C1 or C2.

TS1 corresponds to the transition state in which N3 is nucleophilically attacking the positively charged C1. The forward product from TS1 is IM1, a metastable intermediate located on the potential energy surface, where the C–N bond has formed and the C1–S bond has broken. The free energy barrier from R to IM1 is calculated to be 14.2 kcal/mol.

Once formed, IM1 would be immediately brought to a stable intermediate IM2, which lies below R by 13.7 kcal/mol. Subsequently, IM2 is converted into the precursor of Smiles rearrangement product, P_a , via TS2 with a free energy barrier of 36.1 kcal/mol. In TS2, the C2–S bond is forming and the C–F bond is breaking. From Figure 4, it is clear that the Smiles rearrangement pathway (pathway a) consists of two elementary steps, and the second step is the rate-determining step.

Alternatively, if the reaction proceeds via the direct nucleophilic substitution pathway (pathway b), the transition state involved is TS3, which lies above R by 22.6 kcal/mol. Compared to TS1 involved in pathway a, TS3 is energetically less favorable by 8.4 kcal/mol. Therefore, from an energetic point of view, R prefers to evolve along pathway a, resulting in P_{a} , rather than along pathway b to form P_{b} . The calculated results are in good agreement with the experimental observation that the *ipso*-Smiles rearrangement product is substantially predominant over the ortho position nucleophilic substitution product.

In conclusion, a variety of benzo[b]pyrido[3,2-f][1,4]thiazepin-5(6H)-one and dibenzo[b,f][1,4]thiazepin-11(10H)-one deriv-

The Journal of Organic Chemistry

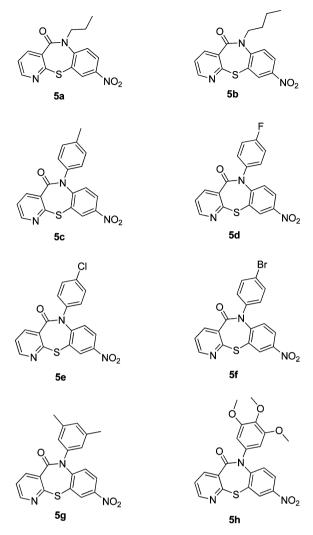


Figure 1. Structures of desired compounds 5a-h.

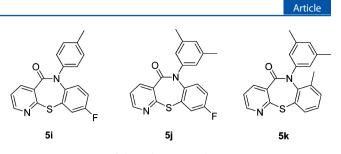


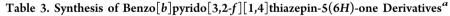
Figure 2. Structures of desired compounds 5i-k.

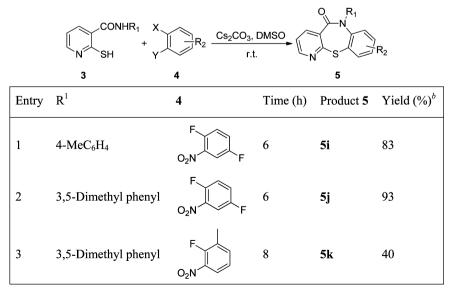
atives were synthesized in good to excellent yields via Smiles rearrangement. The tricyclic systems were constructed by an efficient one-pot transition metal-free process at room temperature. Furthermore, to gain a better understanding of the reaction mechanism, a theoretical study was performed by quantum chemistry calculations. The results show that the Smiles rearrangement pathway has a barrier that is lower by 14.2 kcal/mol⁻¹; thus, it is energetically more favorable than the direct intramolecular nucleophilic substitution pathway. This green and clean synthetic methodology has potential applications in the synthesis of biologically and medicinally relevant compounds.

EXPERIMENTAL SECTION

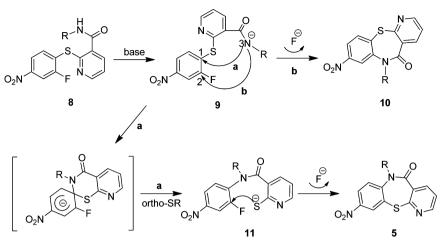
General Procedure. ¹H and ¹³CNMR spectra were recorded with a 300 spectrometer or a 400 spectrometer in CDCl₃. HRMS spectra were recorded on a Q-TOF spectrograph. Compounds 3a-h were prepared according to the literature. Other reagents (Adamas) were commercially available and were used without further purification. All reactions were monitored by thin-layer chromatography (TLC).

Computational Methods. All calculations were conducted using the B3LYP functional^{16,17} combined with the 6-311+(d,p) basis set, as implemented in Gaussian 03.¹⁸ Vibrational analyses at the same level of theory were also conducted to confirm all stationary points as minima (zero imaginary frequencies) or first-order saddle points (one imaginary frequency) and to provide free energies at 298.15 K. Solvent effects have also been taken into account by calculating the singlepoint energies of the geometries obtained in the gas phase by using the simple self-consistent reaction field (SCRF) method¹⁹ based on the polarizable continuum (PCM)²⁰ model. Dimethyl sulfoxide (DMSO)





^aReaction conditions: 2-mercaptonicotinamides 3 (1.0 equiv), substituted benzenes 4 (1.0 equiv), Cs₂CO₃ (3.0 equiv). ^bIsolated yields.



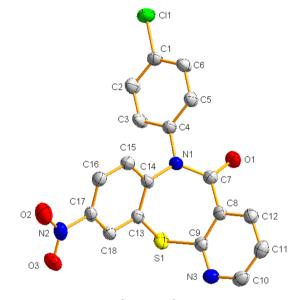


Figure 3. X-ray structure of compound 5e.

was employed as a solvent, corresponding to the experimental conditions.

General Experimental Procedure for 9-Nitro-6-propylbenzo-[b]pyrido[3,2-f][1,4]thiazepin-5(6H)-one (5a). To a solution of DMSO (15 mL) were added 2-mercapto-N-propylnicotinamide 3a (0.20 g, 1.0 mmol), 1,2-difluoro-4-nitrobenzene 4a (0.08 g, 0.5 mmol), and Cs₂CO₃ (0.98 g, 3.0 mmol), and the mixture was stirred for 1 h at room temperature; then another 1,2-difluoro-4-nitrobenzene 4a (0.08 g, 0.5 mmol) was added and the mixture stirred at rt for 4 h. Brine (40 mL) was poured into the solution, and the mixture was extracted with CH_2Cl_2 (3 × 40 mL). The organic layers were combined and dried with anhydrous Na₂SO₄. The product was purified by flash chromatography on silica gel (5:1 hexane/EtOAc). Compound 5a was obtained as a white solid (0.24 g, 0.78 mmol): mp 153-155 °C; ¹H NMR (CDCl₃, 300 MHz) δ 8.61 (d, J = 2.7 Hz, 1H), 8.50 (dd, J = 4.8, 1.8 Hz, 1H), 8.22 (dd, J = 9.0, 2.7 Hz, 1H), 8.02 (dd, J = 7.8, 1.8 Hz, 1H), 7.54 (d, J = 9.0 Hz, 1H), 7.34 (dd, J = 7.5, 4.8 Hz, 1H), 4.65-4.75 (m, 1H), 3.60-3.69 (m, 1H), 1.59-1.78 (m, 2H), 0.96 (t, J = 7.4 Hz, 3H); 13 C NMR (CDCl₃, 75 MHz) δ 166.5, 157.4, 151.3, 148.3, 144.9, 140.1, 135.1, 133.8, 129.5, 126.1, 124.9, 124.0, 53.0, 21.5, 11.4; HRMS calcd for $C_{15}H_{13}N_3O_3S\ [(M\ +\ H)^+]$ 316.0750, found 316.0747.

General Experimental Procedure for 7-Nitro-10-(p-tolyl)dibenzo[b,f][1,4]thiazepin-11(10H)-one (7a). This compound was prepared in the same way as described for **5a** by using 2-mercapto-*N*-(*p*-tolyl)benzamide (0.24 g, 1.0 mmol), 1,2-difluoro-4nitrobenzene **4a** (0.16 g, 1.0 mmol), and Cs₂CO₃ (0.98 g, 3.0 mmol) in DMSO (15 mL) at room temperature. The product was purified by flash chromatography on silica gel (5:1 hexane/EtOAc) to afford 7a (0.34 g, 0.9 mmol) as a white solid: mp 62–64 °C; ¹H NMR (CDCl₃, 400 MHz) δ 8.53 (d, *J* = 2.6 Hz, 1H), 7.97 (dd, *J* = 9, 2.6 Hz, 1H), 7.79–7.81 (m, 1H), 7.51–7.54 (m, 1H), 7.39–7.42 (m, 2H), 7.24–7.29 (m, 4H), 7.13 (d, *J* = 9.0 Hz, 1H), 2.40 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 168.3, 150.2, 144.2, 140.2, 138.3, 138.1, 137.1, 135.7, 131.8, 131.7, 131.3, 130.4, 129.4, 128.4, 128.1, 124.2, 21.2; FT-HRMS (ESI) calcd for C₂₀H₁₄N₂O₃S [(M + H)⁺] 363.0798, found 363.0803.

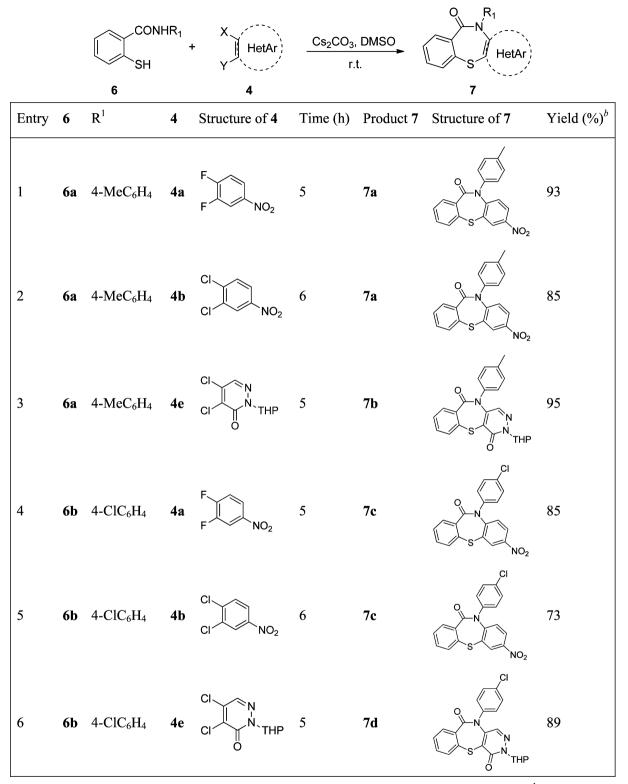
Article

X-ray Crystal Structure Analysis of Compound 5e.²¹ Single crystals of **5e** suitable for X-ray crystal analysis were obtained by recrystallization from a hexane/CH₂Cl₂ mixed solvent. Intensity data were collected at 293 K on an X-ray diffractometer with Mo K α radiation ($\lambda = 0.71073$ Å) and a graphite monochromator. A total of 9276 reflections were measured at a maximal 2 θ angle of 50.0°, of which 3792 were independent reflections ($R_{int} = 0.0500$). The structure was determined by direct methods (SHELXS-97)²² and refined by the full-matrix least-squares method on F^2 (SHELEXL-97).²² The crystal data are as follows: C₁₈H₁₀ClN₃O₃S, $F_W = 383.01$, crystal size of 0.12 mm × 0.10 mm × 0.10 mm, monoclinic, P121/c1, a = 8.784(3) Å, b = 16.217(5) Å, c = 14.313(4) Å, V = 1700.5(9) Å³, Z = 4. The refinement converged to $R_1 = 0.0467$, $wR_2 = 0.1352$ [$I > 2\sigma(I)$], and GOF = 1.111.

6-Butyl-9-nitrobenzo[b]pyrido[**3**,**2**-*f*][**1**,**4**]**thiazepin-5(6***H***)-one** (**5b**): 264 mg (80% yield); pale yellow oil; ¹H NMR (CDCl₃, 300 MHz) δ 8.60 (d, *J* = 1.8 Hz, 1H), 8.49 (dd, *J* = 4.8, 1.2 Hz, 1H), 8.22 (dd, *J* = 8.7, 2.1 Hz, 1H), 8.01 (dd, *J* = 7.8, 1.2 Hz, 1H), 7.55 (d, *J* = 9.0 Hz, 1H), 7.33 (d, *J* = 7.8, 5.1 Hz, 1H), 4.69–4.79 (m, 1H), 3.62–3.71 (m, 1H), 1.58–1.69 (m, 2H), 1.32–1.1.44 (m, 2H), 0.90 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (CDCl₃, 75 MHz) δ 166.4, 157.4, 151.3, 148.3, 144.9, 140.1, 135.1, 133.8, 129.4, 126.1, 124.9, 124.0, 51.0, 30.2, 20.1, 13.7; FT-HRMS (ESI) calcd for C₁₆H₁₅N₃O₃S [(M + H)⁺] 330.0907, found 330.0933.

9-Nitro-6-(*p***-tolyl)benzo[***b***]pyrido[3,2-***f***][1,4]thiazepin-5(6***H***)one (5c): 326 mg (90% yield); white solid; mp 185–187 °C; ¹H NMR (CDCl₃, 300 MHz) \delta 8.64 (d,** *J* **= 2.7 Hz, 1H), 8.55 (dd,** *J* **= 4.8, 1.8 Hz, 1H), 8.12 (dd,** *J* **= 7.8, 1.8 Hz, 1H), 8.02 (dd,** *J* **= 9.0, 2.7 Hz, 1H), 7.39 (dd,** *J* **= 7.8, 4.8 Hz, 1H), 7.24–7.32 (m, 5H), 7.19 (d,** *J* **= 9.0 Hz, 1H), 2.41 (s, 3H); ¹³C NMR (CDCl₃, 75 MHz) \delta 166.5, 157.0, 151.7, 149.1, 144.6, 140.2, 139.5, 138.7, 134.1, 133.2, 130.4, 129.4, 128.1, 124.5, 124.1, 21.2; FT-HRMS (ESI) calcd for C₁₉H₁₃N₃O₃S [(M + H)⁺] 364.0750, found 364.0766.**

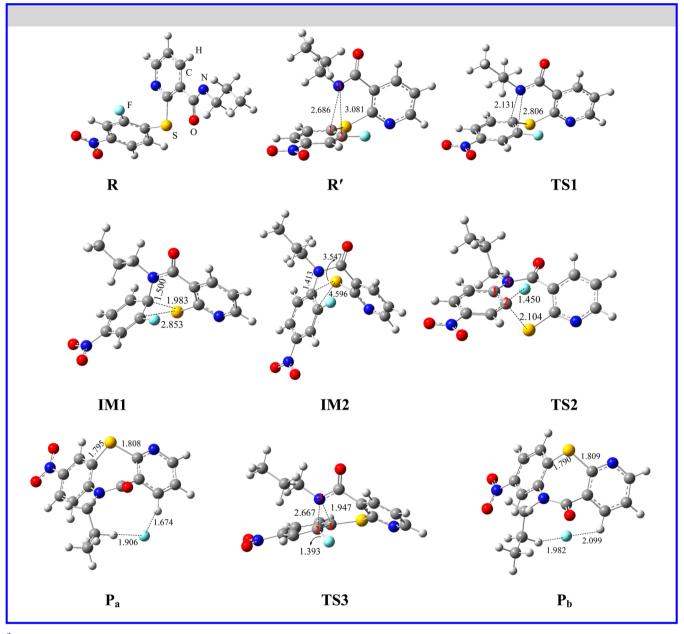
6-(4-Fluorophenyl)-9-nitrobenzo[b]pyrido[3,2-f][1,4]thiazepin-5(6H)-one (5d): 322 mg (88% yield); white solid; mp 148–150 °C; ¹H NMR (CDCl₃, 300 MHz) δ 8.65 (d, J = 2.4 Hz, 1H), Table 4. Synthesis of 1,4-Thiazepin-5(4H)-one Derivatives^a



"Reaction conditions: 2-mercaptonicotinamides 3 (1.0 equiv), substituted benzenes 4 (1.0 equiv), Cs₂CO₃ (3.0 equiv). ^bIsolated yields.

8.56 (d, J = 3.3 Hz, 1H), 8.13 (d, J = 1.2 Hz, 1H), 8.05 (dd, J = 9.0, 2.7 Hz, 1H), 7.36–7.42 (m, 3H), 7.15–7.22 (m, 3H); ¹³C NMR (CDCl₃, 75 MHz) δ 166.5, 162.0 (${}^{1}J_{C,F}$ = 248 Hz), 157.0, 151.9, 148.8, 144.8, 140.3, 137.9 (${}^{4}J_{C,F}$ = 3 Hz), 133.5 (${}^{2}J_{C,F}$ = 19 Hz), 130.2 (${}^{3}J_{C,F}$ = 9 Hz), 129.5, 128.0, 124.7, 124.2, 117.0, 116.6; FT-HRMS (ESI) calcd for C₁₈H₁₀FN₃O₃S [(M + H)⁺] 368.0500, found 368.0516.

6-(4-Chlorophenyl)-9-nitrobenzo[*b*]**pyrido**[**3**,**2**-*f*][**1**,**4**]**thiazepin-5(6***H***)-one (5e):** 363 mg (95% yield); white solid; mp 195–196 °C; ¹H NMR (CDCl₃, 300 MHz) δ 8.65 (d, *J* = 2.7 Hz, 1H), 8.56 (dd, *J* = 4.8, 1.8 Hz, 1H), 8.11 (dd, *J* = 7.8, 1.8 Hz, 1H), 8.05 (dd, *J* = 9, 2.7 Hz, 1H), 7.45–7.50 (m, 2H), 7.39 (dd, *J* = 7.8, 4.8 Hz, 1H), 7.31–7.36 (m, 2H), 7.16 (d, *J* = 9.0 Hz, 1H); ¹³C NMR (CDCl₃, 75 MHz) δ 166.3, 157.0, 151.9, 148.6, 144.9, 140.4, 140.3, 134.4, 133.7, Table 5. Optimized Structures for Transition States, Intermediates, and Products Involved along Pathways a and b Shown in Scheme 3^a



^aDistances are in angstroms.

133.6, 130.0, 129.7, 129.5, 128.1, 124.7, 124.2; FT-HRMS (ESI) calcd for $C_{18}H_{10}ClN_3O_3S$ [(M + H)⁺] 384.0204, found 384.0214.

6-(**4**-Bromophenyl)-9-nitrobenzo[*b*]pyrido[3,2-*f*][1,4]thiazepin-5(6*H*)-one (5f): 388 mg (91% yield); white solid; mp 216–218 °C; ¹H NMR (CDCl₃, 300 MHz) δ 8.65 (d, *J* = 2.7 Hz, 1H), 8.56 (dd, *J* = 4.8, 1.8 Hz, 1H), 8.11 (dd, *J* = 7.8, 1.8 Hz, 1H), 8.05 (dd, *J* = 9.0, 2.7 Hz, 1H), 7.60–7.65 (m, 2H), 7.39 (dd, *J* = 7.8, 4.8 Hz, 1H), 7.25–7.29 (m, 2H), 7.15 (d, *J* = 9.0 Hz, 1H); ¹³C NMR (CDCl₃, 75 MHz) δ 166.2, 157.0, 151.9, 148.5, 144.9, 141.0, 140.3, 133.7, 133.0, 130.0, 129.5, 128.1, 124.7, 124.2, 122.5; FT-HRMS (ESI) calcd for C₁₈H₁₀BrN₃O₃S [(M + H)⁺] 427.9699, found 427.9673.

6-(3,5-Dimethylphenyl)-9-nitrobenzo[*b*]**pyrido**[**3,2-***f*][**1,4**]**thiazepin-5(6H)-one (5g):** 358 mg (95% yield); white solid; mp none; ¹H NMR (CDCl₃, 400 MHz) δ 8.64 (d, *J* = 2.4 Hz, 1H), 8.55 (d, *J* = 1.4 Hz, 1H), 8.10 (d, *J* = 7.7 Hz, 1H), 8.02 (dd, *J* = 9.0, 2,4 Hz, 1H), 7.38 (dd, *J* = 7.7, 2.9 Hz, 1H), 7.22 (d, *J* = 9.0 Hz, 1H), 7.04 (s, 1H), 6.99 (s, 2H), 2.36 (s, 6H); ¹³C NMR (CDCl₃, 100 MHz) δ 166.4, 157.0, 151.6, 149.0, 144.6, 141.9, 140.1, 139.7, 134.2, 133.1, 130.4, 129.4, 128.0, 126.0, 124.5, 124.1, 21.2; FT-HRMS (ESI) calcd for $C_{20}H_{15}N_3O_3S$ [(M + H)⁺] 378.0907, found 378.0919.

9-Nitro-6-(3,4,5-trimethoxyphenyl)benzo[b]pyrido[3,2-f] [1,4]thiazepin-5(6H)-one (5h): 395 mg (90% yield); white solid; mp 228–230 °C; ¹H NMR (CDCl₃, 300 MHz) δ 8.65 (d, *J* = 2.7 Hz, 1H), 8.56 (dd, *J* = 4.8, 1.8 Hz, 1H), 8.12 (dd, *J* = 7.8, 1.8 Hz, 1H), 8.07 (dd, *J* = 9.0, 2.7 Hz, 1H), 7.39 (dd, *J* = 7.8, 4.8 Hz, 1H), 7.30 (d, *J* = 9.0 Hz, 1H), 6.58 (s, 2H), 3.89 (s, 3H), 3.87 (s, 6H); ¹³C NMR (CDCl₃, 75 MHz) δ 166.5, 156.9, 154.0, 151.8, 148.8, 144.7, 140.2, 138.3, 137.4, 134.0, 132.9, 129.5, 127.7, 124.7, 124.2, 106.0, 60.9, 56.4; FT-HRMS (ESI) calcd for C₂₁H₁₇N₃O₆S [(M + H)⁺] 440.0872, found 440.0853. **9-Fluoro-6-(***p***-tolyl)benzo[***b***]pyrido[3,2-***f***][1,4]thiazepin-5(6***H***)-one (5i): 278 mg (83% yield); white solid; mp 152–153 °C; ¹H NMR (CDCl₃, 300 MHz) \delta 8.51 (dd,** *J* **= 4.8, 1.8 Hz, 1H), 8.12 (dd,** *J* **= 7.8, 1.8 Hz, 1H), 7.46 (dd,** *J* **= 7.8, 3 Hz, 1H), 7.35 (dd,** *J* **= 7.8, 4.8 Hz, 1H), 7.26 (s, 5H), 7.06 (dd,** *J* **= 9.0, 5.1 Hz, 1H), 6.87–**

The Journal of Organic Chemistry

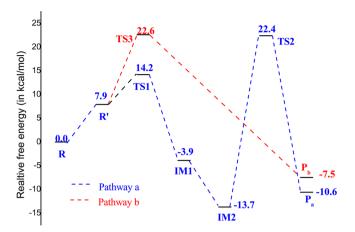


Figure 4. Calculated relative free energy profiles along the *ipso*-Smiles rearrangement pathway (pathway a) and the direct nucleophilic substitution pathway (pathway b).

6.94 (m, 1H), 2.39 (m, 3H); ¹³C NMR (CDCl₃, 75 MHz) δ 172.3, 166.7, 162.8 (${}^{1}J_{C,F}$ = 232 Hz), 158.1, 157.8, 156.1, 150.9, 140.1 (${}^{3}J_{C,F}$ = 9 Hz), 140.0, 137.8, 134.5, 130.1, 129.1 (${}^{3}J_{C,F}$ = 8 Hz), 127.8, 123.8, 120.4 (${}^{2}J_{C,F}$ = 22 Hz), 117.1 (${}^{2}J_{C,F}$ = 22 Hz), 21.2; ¹⁹F NMR (CDCl₃, 282 MHz) δ –114.23; FT-HRMS (ESI) calcd for C₁₉H₁₃FN₂OS [(M + H)⁺] 337.0766, found 337.0790.

6-(**3**,**5**-Dimethylphenyl)-9-fluorobenzo[*b*]pyrido[**3**,**2**-*f*][**1**,**4**]-thiazepin-5(6*H*)-one (**5**): 315 mg (93% yield); white solid; mp 148–150 °C; ¹H NMR (CDCl₃, 300 MHz) δ 8.52 (dd, *J* = 4.8, 1.5 Hz, 1H), 8.12 (d, *J* = 4.8 Hz, 1H), 7.46 (dd, *J* = 7.8, 2.7 Hz, 1H), 7.36 (dd, *J* = 7.8, 4.8 Hz, 1H), 7.10 (dd, *J* = 9.0, 5.1 Hz, 1H), 7.00 (s, 3H), 6.88–6.95 (m, 1H), 2.35 (s, 6H); ¹³C NMR (CDCl₃, 75 MHz) δ 165.6, 157.0,156.8, 149.8, 141.5, 139.2, 138.9 (⁴*J*_{C,F} = 4 Hz), 138.3, 133.6, 133.1 (³*J*_{C,F} = 9 Hz), 128.7, 128.1 (³*J*_{C,F} = 9 Hz), 124.7, 122.9, 119.5 (²*J*_{C,F} = 23 Hz), 116.1 (²*J*_{C,F} = 22 Hz), 20.3; ¹⁹F NMR (CDCl₃, 282 MHz) δ –114.36; FT-HRMS (ESI) calcd for C₂₀H₁₅FN₂OS [(M + H)⁺] 351.0962, found 351.0972.

6-(3,5-Dimethylphenyl)-7-methylbenzo[*b*]**pyrido**[**3,2-***f*][**1,4**]-**thiazepin-5(6***H*)-**one** (**5k**): 138 mg (40% yield); white solid; mp 190–192 °C; ¹H NMR (CDCl₃, 300 MHz) δ 8.43 (dd, *J* = 4.8, 1.8 Hz, 1H), 8.06 (dd, *J* = 7.5, 1.8 Hz, 1H), 7.68 (dd, *J* = 7.2, 2.4 Hz, 1H), 7.30 (dd, *J* = 7.8, 4.8 Hz, 1H), 7.08–7.15 (m, 2H), 6.98 (s, 2H), 6.86 (s, 1H), 2.30 (s, 6H), 2.00 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 166.8, 150.0, 141.4, 140.8, 139.5, 137.9, 137.3, 136.8, 135.1, 132.9, 132.1, 128.2, 127.4, 124.7, 123.6, 21.4, 20.0; FT-HRMS (ESI) calcd for C₂₁H₁₈N₂OS [(M + H)⁺] 347.1213, found 347.1224.

9-(Tetrahydro-2*H***-pyran-2-yl)-6-(***p***-tolyl)pyridazino[4,5-***b***]pyrido[3,2-***f***][1,4]thiazepine-5,10(6***H***,9***H***)-dione (7***b***): 398 mg (95% yield); white crystal; mp 98–100 °C; ¹H NMR (CDCl₃, 300 MHz) \delta 7.77 (m, 1H), 7.52–7.57 (m, 1H), 7.39–7.46 (m, 2H), 7.35 (s, 1H), 7.22–7.31 (m, 4H), 5.99 (dd,** *J* **= 10.5, 1.8 Hz, 1H), 4.07 (d,** *J* **= 11.4 Hz, 1H), 3.68–3.76 (m, 1H), 2.40 (s, 3H), 1.92–2.09 (m, 2H), 1.51–1.76 (m, 4H); ¹³C NMR (CDCl₃, 75 MHz) \delta 168.7, 145.3, 139.2, 138.1, 137.0, 136.9, 134.4, 132.4, 132.0, 131.9, 130.5, 129.3, 128.3, 83.6, 68.8, 28.6, 24.8, 22.7, 21.2; FT-HRMS (ESI) calcd for C₂₃H₂₁N₃O₃S [(M + H)⁺] 420.1376, found 420.1382.**

10-(4-Chlorophenyl)-7-nitrodibenzo[*b*,*f*][1,4]thiazepin-11-(10*H*)-one (7c): 324 mg (85% yield); white solid; mp 98–100 °C; ¹H NMR (CDCl₃, 400 MHz) δ 8.54 (d, *J* = 2.6 Hz, 1H), 8.00 (dd, *J* = 8.9, 2.6 Hz, 1H), 7.78–7.81 (m, 1H), 7.53–7.55 (m, 1H), 7.40–7.46 (m, 4H), 7.33 (d, *J* = 8.7 Hz, 1H), 7.10 (d, *J* = 9.0 Hz, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 168.1, 149.7, 144.5, 141.1, 137.6, 137.1, 136.2, 134.1, 131.9, 131.8, 131.4, 130.0, 129.9, 129.5, 128.6, 128.1, 124.4; FT-HRMS (ESI) calcd for C₁₉H₁₁ClN₂O₃S [(M + H)⁺] 383.0252, found 383.0253.

11-(4-Chlorophenyl)-3-(tetrahydro-2*H*-pyran-2-yl)benzo[*f*]pyridazino[4,5-*b*][1,4]thiazepine-4,10(3*H*,11*H*)-dione (7d): 390 mg (89% yield); white crystal; mp 138–140 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.79 (d, *J* = 4.9 Hz, 1H), 7.55 (d, *J* = 3.7 Hz, 1H), 7.43– 7.52 (m, 4H), 5.98 (d, J = 10.2 Hz, 1H), 4.07 (d, J = 11.4 Hz, 1H), 7.31 (d, J = 4.0 Hz, 3H) 1.98–2.05 (m, 2H), 1.52–1.70 (m, 4H); ¹³C NMR (CDCl₃, 100 MHz) δ 168.5, 158.4, 144.9, 138.1, 137.7, 137.0, 135.0, 134.2, 132.6, 132.3, 132.1, 130.1, 130.0, 129.4. 83.8, 68.9, 28.7, 24.8, 22.7; FT-HRMS (ESI) calcd for C₂₂H₁₈ClN₃O₃S [(M + H)⁺] 440.0830, found 440.0864.

Article

ASSOCIATED CONTENT

S Supporting Information

Copies of ¹H and ¹³C NMR spectra of all compounds, X-ray data of **5e** in CIF format, and Cartesian coordinates and absolute energies for all structures involved in theoretical calculations. This material is available free of charge via the Internet at http://pubs.acs.org.

AUTHOR INFORMATION

Corresponding Authors

*E-mail: chenma@sdu.edu.cn.

*E-mail: zhangdj@sdu.edu.cn.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

We are grateful to the National Science Foundation of China (Grants 21172131 and 21273131) and the State Key Laboratory of Bioactive Substance and Function of Natural Medicines, Institute of Materia Medica, Chinese Academy of Medical Sciences, and Peking Union Medical College (GTZK201405) for financial support of this research.

REFERENCES

 (a) Masumiya, H.; Tanaka, H.; Shigenobu, K. Eur. J. Pharmacol.
 1997, 335, 15–21.
 (b) Watanabe, T.; Kalasz, H.; Yabana, H.; Kuniyasu, A.; Mershon, J.; Itagaki, K.; Vaghy, P. L.; Naito, K.; Nakayama, H.; Schwartz, A. FEBS Lett. 1993, 334, 261–264.
 (c) Bariwal, J. B.; Upadhyay, K. D.; Manvar, A. T.; Trivedi, J. C.; Singh, J. S.; Jain, K. S.; Shah, A. K. Eur. J. Med. Chem. 2008, 43, 2279– 2290.
 (d) Wajima, T.; Fukumura, K.; Yano, Y.; Oguma, T. J. Pharm. Sci. 2003, 92, 2427–2440.
 (e) Beaucage, P.; Massicotte, J.; Boileau, J. F.; Dumont, L. J. Cardiovasc. Pharmacol. 2003, 42, 142–150.

(2) (a) Geller, V.; Gorzaltsan, I.; Shleifer, T.; Belmaker, R. H.; Bersudsky, Y. Schizophr. Res. Treat. 2005, 80, 343–347. (b) Kurita, M.; Holloway, T.; García-Bea, A.; Kozlenkov, A.; Friedman, A. K.; Moreno, J. L.; Heshmati, M.; Golden, S. A.; Kennedy, P. J.; Takahashi, N.; Dietz, D. M.; Mocci, G.; Gabilondo, A. M.; Hanks, J.; Umali, A.; Callado, L. F.; Gallitano, A. L.; Neve, R. L.; Shen, L.; Buxbaum, J. D.; Han, M. H.; Nestler, E. J.; Meana, J. J.; Russo, S. J.; González-Maeso, J. Nat. Neurosci. 2012, 15, 1245–1254. (c) Kapur, S.; Seeman, P. Am. J. Psychiatry 2001, 158, 360. (d) Richelson, E.; Souder, T. Life Sci. 2000, 68, 29–39. (e) Seminara, G.; Trassari, V.; Prestifilippo, N.; Chiavetta, R.; Calandra, C. Minerva Psichiatrica 1993, 34, 95–99.

(3) Pettersson, H.; Bülow, A.; Ek, F.; Jensen, F. E.; Otteson, L. K.; Fejzic, A.; Ma, J.-N.; Del Tredici, A. L.; Currier, E. A.; Gardell, L. R.; Tabatabaei, A.; Craig, D.; McFarland, K.; Ott, T. R.; Piu, F.; Burstein, E. S.; Olsson, R. J. Med. Chem. **2009**, *52*, 1975–1982.

(4) Binaschi, M.; Boldetti, A.; Gianni, M.; Maggi, C. A.; Gensini, M.; Bigioni, M.; Parlani, M.; Giolitti, A.; Fratelli, M.; Valli, C.; Terao, M.; Garattini, E. ACS Med. Chem. Lett. **2010**, *1*, 411–415.

(5) (a) Kumar, M.; Sharma, K.; Fogla, A. K.; Sharma, K.; Rathore, M. Res. Chem. Intermed. 2013, 39, 2555–2564. (b) Anisetti, R.; Reddy, M. S. J. Sulfur Chem. 2012, 33, 363–372. (c) Sayed, G. H.; Sayed, M. A.; Mahmoud, M. R.; Shaaban, S. S. Egypt. J. Chem. 2002, 45, 767–776. (d) Patel, R. N.; Nimavat, K. S.; Vyas, K. B.; Patel, P. V. J. Chem. Pharm. Res. 2011, 3, 409–415.

(6) Frolov, E. B.; Lakner, F. J.; Khvat, A. V.; Ivachtchenko, A. V. Tetrahedron Lett. 2004, 45, 4693–4696.

The Journal of Organic Chemistry

(7) (a) Robl, J. A.; Sun, C. Q.; Stevenson, J.; Ryono, D. E.; Simpkins, L. M.; Cimarusti, M. P.; Dejneka, T. W.; Slusarchyk, A.; Chao, S.; Stratton, L.; Misra, R. N.; Bednarz, M. S.; Asaad, M. M.; Cheung, H. S.; Abboa-Offei, B. E.; Smith, P. L.; Mathers, P. D.; Fox, M.; Schaeffer, T. R.; Seymour, A. A.; Trippodo, N. C. J. Med. Chem. **1997**, 40, 1570– 1577. (b) Barbaro, R.; Betti, L.; Botta, M.; Corelli, F.; Giannaccini, G.; Maccari, L.; Manetti, F.; Strappaghetti, G.; Corsano, S. J. Med. Chem. **2001**, 44, 2118–2132. (c) Oda, T.; Tijima, Y.; Sada, T.; Nishino, H.; Oizumi, K.; Koike, H. Biochem. Biophys. Res. Commun. **1988**, 152, 456–462.

(8) Katrusiak, B.; Unlu, S.; Banoglu, E.; Kupeli, E.; Yesilada, E.; Sahin, M. F. Arch. Pharm. Pharm. Med. Chem. **2003**, 336, 406–412.

(9) Libgeois, J. F.; Rogistert, F. A.; Bruhwyler, J.; Damas, J.; Nguyen, T. P.; Inarejos, M. O.; Eric, M. G. J. Med. Chem. **1994**, 37, 519–525. (10) Fu, R.; Xiu, X.; Dang, Q.; Bai, X. J. Org. Chem. **2005**, 70, 10810–

(10) Fu, K., Xid, X., Dang, Q., Bai, K. J. Org. Chem. 2003, 70, 10810– 10816. (11) Binaschi, M.; Boldetti, A.; Gianni, M.; Maggi, C. A.; Gensini, M.;

(11) Binaschi, M.; Boldetti, A.; Gianni, M.; Maggi, C. A.; Gensini, M.; Bigioni, M.; Parlani, M.; Giolitti, A.; Fratelli, M.; Valli, C.; Terao, M.; Garattini, E. ACS Med. Chem. Lett. **2010**, *1*, 411–415.

(12) Ottesen, L. K.; Fredrik, E.; Roger, O. Org. Lett. 2006, 8, 1771–1773.

(13) Panda, N.; Jena, A. K.; Mohapatra, S. *Appl. Catal., A* **2012**, 433–434, 258–264.

(14) (a) Huang, A. P.; Chen, Y. M.; Zhou, Y. G.; Guo, W.; Wu, X. D.; Ma, C. Org. Lett. 2013, 15, 5480–5483. (b) Zhao, Y. M.; Wu, Y. M.; Jia, J.; Zhang, D. J.; Ma, C. J. Org. Chem. 2012, 77, 8501–8506.
(c) Yang, B. C.; Niu, X. Y.; Huang, Z. X.; Zhao, C. H.; Liu, Y.; Ma, C. Tetrahedron 2013, 69, 8250–8254. (d) Yang, B. C.; Huang, Z. X.; Guan, H. G.; Niu, X. Y.; Li, Y. Q.; Fang, S.; Ma, C. Tetrahedron Lett. 2013, 54, 5994–5997. (e) Niu, X. Y.; Yang, B. C.; Li, Y. Q.; Fang, S.; Huang, Z. X.; Xie, C. X.; Ma, C. Org. Biomol. Chem. 2013, 11, 4102– 4108. (f) Liu, Y. L.; Zhan, C. J.; Yang, B. C.; Cao, X. Q.; Ma, C. Synlett 2013, 45, 111–117. (g) Liu, Y. L.; Chu, C. X.; Huang, A. P.; Zhan, C. J.; Ma, Y.; Ma, C. ACS Comb. Sci. 2011, 13, 547–553.

(15) Kohn, W.; Becke, A. D.; Parr, R. G. J. Phys. Chem. 1996, 100, 12974–12980.

(16) Lee, C.; Yang, W.; Parr, G. Phys. Rev. B 1988, 37, 785-794.

(17) Becke, A. D. J. Chem. Phys. 1993, 98, 5648-5652.

(18) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Montgomery, J. A., Jr.; Vreven, T.; Kudin, K. N.; Burant, J. C.; Millam, J. M.; Iyengar, S. S.; Tomasi, J.; Barone, V.; Mennucci, B.; Cossi, M.; Scalmani, G.; Rega, N.; Petersson, G. A.; Nakatsuji, H.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Klene, M.; Li, X.; Knox, J. E.; Hratchian, H. P.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Ayala, P. Y.; Morokuma, K.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Zakrzewski, V. G.; Dapprich, S.; Daniels, A. D.; Strain, M. C.; Farkas, O.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Ortiz, J. V.; Cui, Q.; Baboul, A. G.; Clifford, S.; Cioslowski, J.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen, W.; Wong, M. W.; Gonzalez, C.; Pople, J. A. Gaussian 03; Gaussian, Inc.: Wallingford, CT, 2004.

(19) Tapia, O. J. Math. Chem. 1992, 10, 139-181.

(20) Barone, V.; Cossi, M.; Tomasi, J. J. Comput. Chem. 1998, 19, 404-407.

(21) For the crystal data of **5e**, see the Supporting Information. CCDC 994893 (**5e**) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crysallographic Data Centre via www.ccdc.cam.ac.uk/ data request/cif.

(22) Sheldrick, G. M. SHELX-97, Program for the Refinement of Crystal Structure; University of Göttingen: Göttingen, Germany, 1997.